

### Supplementary Discussion

Our study brings to light a basic distinction in how EGFs and Upd cytokines are deployed for steady-state turnover versus injury repair. During steady-state turnover, apoptotic induction of *rho* is strictly cell autonomous (Fig. 4d-f) because E-cad-dependent activation of p120 and Arm is confined to the dying enterocyte. Of note, neither physiological apoptosis nor *E-cad* depletion activates *upd* cytokines, including the cardinal injury signal, *upd3* (Extended Data Fig. 6a-d, l). Autonomous induction of *rho* and consequent release of EGFs by the apoptotic cell confines mitogenic signaling to the precise time and place that division is needed, as appropriate for zero-sum cell replacement.

By contrast, others have shown that tissue-wide injury invokes a separate, non-cell autonomous pathway: Upon pan-enterocyte death or pathogenic infection, damaged enterocytes upregulate *upd3*, and Upd3 in turn activates enteroblasts and visceral muscle to upregulate *rho* and *EGFs*<sup>15,16,20,21</sup>. This non-autonomous relay causes EGFs to be released in a widespread, indiscriminate manner, as appropriate for an emergency response.

Underscoring these distinctions, enterocyte *upd3* is required for repair<sup>6,16,20,21</sup> but not homeostasis (Extended Data Figs. 6a-d, k and 7i), whereas enterocyte *rho* is required for homeostasis (Fig. 4k, Extended Data Fig. 8) but not repair<sup>16</sup>. These contrasts imply that dying cells signal differently in injury and steady-state contexts, possibly reflecting loss of the intestinal barrier or inefficient clearance of cell corpses following extensive damage. Thus, steady-state turnover is not the repair of one-cell ‘mini-wounds’, but rather an independent, parallel mechanism.

Supplementary Table 1   Experimental genotypes in each figure				
Figure	Panels	Genotype		
Figure 1	Fig. 1a-e	<i>w; esgGAL4, tubGAL80ts, UAS-GFP; UAS-flp, act&lt;CD2&lt;GAL4</i>		
	Fig. 1g-i	<i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-his2A:RFP</i> <i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-p35</i>		
	Fig. 1j-k	<i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP</i>		
		<i>w; mexGAL4, tubGAL80ts; UAS-p35</i> <i>w; mexGAL4, tubGAL80ts; UAS-diap1</i>		
Figure 2	Fig. 2a	<i>y, w; shg[mTomato]</i>		
	Fig. 2b-g	<i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-his2A:RFP</i> <i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-p35</i> <i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-p35, UAS-E-cad RNAi</i> <i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-E-cad</i>		
		Fig. 2h-j	<i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-E-cad RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35</i>	
			Fig. 2k	<i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-ed RNAi</i> <i>hsflp; X-15-29, tubGAL80ts/X-15-33, esgGAL4; UAS-his2A:RFP</i> <i>hsflp; X-15-29, tubGAL80ts/X-15-33, esgGAL4; UAS-E-cad</i> <i>hsflp; X-15-29, tubGAL80ts/X-15-33, esgGAL4; UAS-E-cad RNAi</i>
				Fig. 3a, g
	Figure 3	Fig. 3b-e	<i>w; mexGAL4, tubGAL80ts; UAS-E-cad</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi</i> <i>w; Egfr<sup>tsla</sup>/Egfr<sup>f24</sup>; mexGAL4 TM2/tubGAL80ts, UAS-E-cad RNAi</i>	
		Fig. 3f	<i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-p35, UAS-E-cad RNAi</i>	
		Fig. 3g	<i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi, UAS-spi RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi, UAS-krn RNAi</i>	
			Fig. 3i-k	<i>w; mexGAL4, tubGAL80ts, UAS-spi RNAi; UAS-p35, UAS-E-cad RNAi, UAS-krn RNAi</i> <i>w UAS-CD8:GFP hsflp; tubGAL4/UAS-E-cad RNAi; FRT82 tubGAL80/FRT82</i> <i>w UAS-CD8:GFP hsflp; tubGAL4; FRT82 tubGAL80/FRT82</i>
				Fig. 4a
Figure 4		Fig. 4a-b	<i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi</i>	
		Fig. 4c	<i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, UAS-rho RNAi</i>	
		Fig. 4d-f	<i>rho<sup>X81</sup> (rho-lacZ)</i>	
		Fig. 4g-k	<i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi, rho RNAi</i>	
			Fig. 4k	<i>w; mexGAL4, tubGAL80ts, UAS-rho; UAS-p35</i>

**Supplementary Table 1 (continued) | Experimental genotypes in each figure**

Figure	Panels	Genotype	
<b>Figure 4</b> (continued)	Fig. 4k	<i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi, UAS-arm RNAi</i>	
		<i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-E-cad RNAi, UAS-p120 RNAi</i>	
		<i>w; mexGAL4, tubGAL80ts, UAS-arm RNAi; UAS-p35, UAS-E-cad RNAi, UAS-p120 RNAi</i>	
		<i>UAS-arm<sup>S10</sup>; mexGAL4, tubGAL80ts; UAS-p35</i>	
		<i>w; mexGAL4, tubGAL80ts; UAS-p35, UAS-p120</i>	
		<i>UAS-arm<sup>S10</sup>; mexGAL4, tubGAL80ts; UAS-p35, UAS-p120</i>	
<b>ED Figure 1</b>	ED Fig. 1f-g	<i>w; esgGAL4, tubGAL80ts, UAS-GFP; UAS-flp, act&lt;CD2&lt;GAL4</i>	
<b>ED Figure 2</b>	ED Fig. 2b	<i>hsflp; X-15-29, tubGAL80ts/X-15-33, mexGAL4; UAS-his2A:RFP</i>	
<b>ED Figure 3</b>	ED Fig. 3a	See corresponding “Total Cells (R4ab)” panels – Figs. 1k, 2h, 3g, 4k	
	ED Fig. 3b	See corresponding “dpErk” panels – Figs. 3, 4, Extended Data Fig. 7	
<b>ED Figure 4</b>	ED Fig. 4a-j	<i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP</i>	
		<i>w; mexGAL4, tubGAL80ts; UAS-p35</i>	
		<i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi</i>	
<b>ED Figure 5</b>	ED Fig. 5a-c	<i>w UAS-CD8:GFP hsflp; tubGAL4; FRT82 tubGAL80/FRT82</i> <i>w UAS-CD8:GFP hsflp; tubGAL4/UAS-E-cad RNAi; FRT82 tubGAL80/FRT82</i>	
	ED Fig. 5d-f	<i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi</i>	
<b>ED Figure 6</b>	ED Fig. 6a	<i>w; mexGAL4, tubGAL80ts; UAS-CD4:GFP (qPCR reference)</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi</i>	
	ED Fig. 6b-d	<i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP, upd3.1-lacZ</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, upd3.1-lacZ</i>	
		ED Fig. 6e-g	<i>w; mexGAL4, tubGAL80ts, 10XSTAT-GFP; UAS-his2A:RFP</i> <i>w; mexGAL4, tubGAL80ts, 10XSTAT-GFP; UAS-E-cad RNAi</i>
	ED Fig. 6h-j	<i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP, cycE-lacZ</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, cycE-lacZ</i>	
		ED Fig. 6k	<i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, Su(H)-lacZ</i>
		ED Fig. 6l	<i>w; upd3.1-lacZ</i>
<b>ED Figure 7</b>	ED Fig. 7a-b	<i>w; mexGAL4, tubGAL80ts; UAS-CD4:GFP (qPCR reference)</i>	
	ED Fig. 7a	<i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, UAS-groucho</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, UAS-puc2A</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, UAS-yki RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, UAS-arm RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, UAS-p120 RNAi</i>	
		ED Fig. 7b	<i>UAS-arm<sup>S10</sup>; mexGAL4, tubGAL80ts</i> <i>w; mexGAL4, tubGAL80ts; UAS-p120</i>
			ED Fig. 7c-f

ED = Extended Data

**Supplementary Table 1 (continued) | Experimental genotypes in each figure**

Figure	Panels	Genotype
<b>ED Figure 7</b> (continued)	ED Fig. 7g-l	See genotypes in Extended Data Fig. 7a <i>w; mexGAL4, tubGAL80ts; UAS-E-cad RNAi, UAS-upd3 RNAi</i> <i>w; mexGAL4, tubGAL80ts, UAS-arm RNAi; UAS-E-cad RNAi, UAS-p120 RNAi</i>
	ED Fig. 7m-n, s	<i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP</i> <i>w; mexGAL4, tubGAL80ts, UAS-rho</i>
	ED Fig. 7o-r	Same as Extended Data Figure 7b <i>w; mexGAL4, tubGAL80ts; UAS-rho RNAi</i> <i>UAS-arm<sup>S10</sup>; mexGAL4, tubGAL80ts; UAS-p120</i>
	<b>ED Figure 8</b>	ED Fig. 8a-d <i>w; mexGAL4, tubGAL80ts; UAS-his2A:RFP</i> <i>w; mexGAL4, tubGAL80ts, UAS-rho RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-arm RNAi</i> <i>w; mexGAL4, tubGAL80ts; UAS-p120 RNAi</i>

ED = Extended Data

**Supplementary Table 2 | List of qPCR primers used in this study**

Target	Forward Primer	Reverse Primer
<i>vein</i>	GAACGCAGAGGTCACGAAGA	GAGCGCACTATTAGCTCGGA
<i>spitz</i>	CGCCCAAGAATGAAAGAGAG	AGGTATGCTGCTGGTGAAC
<i>keren</i>	CGTGTGGCAACAACAAGT	TGTGGCAATGCAGTTTAAGG
<i>egfr</i>	TGCATCGGCACTAAATCTCGG	GGAAGCTGAGGTCCAAATTCTC
<i>argos</i>	TGCTGTTGGGTGAATTCAGG	CGACTGGTCCAGATGATCCA
<i>star</i>	AGCCCAGTCCTTCAAACCC	CCACAGTCTTTGGTTGGTTGC
<i>rhomboid</i>	GAGCACATCTACATGCAACGC	GGAGATCACTAGGATGAACCAGG
<i>frizzled-3</i>	TCTTGTGCCCGCAAACCTTTA	CCTAGAATGAGGGTCTCAGACG
<i>senseless</i>	GATCGTGACTTTGCCTTGACG	CCTGATAGTCCTGCTTGCTGT
<i>expanded</i>	GATGCTGGACACCGAACTCT	CTTGCTCTCGGGATCTGC
<i>diap1</i>	GAAAAAGAGAAAAGCCGTCAAGT	TGTTTGCCTGACTCTTAATTTCTTC
<i>pointed</i>	CTACGAGAAGCTGAGTCGCG	TATCGTTTGCCTGCCGTCTT
<i>cyclin-E</i>	ACAAATTTGGCCTGGGACTA	GGCCATAAGCACTTCGTCA
<i>unpaired-1</i>	CCTACTCGTCCTGCTCCTTG	TGCGATAGTCGATCCAGTTG
<i>unpaired-2</i>	GAGGGCAGCTACGACAGTG	GGAGAAGAGTCGCAGTTGT
<i>unpaired-3</i>	AAATTCGACAAAGTCGCCTG	TTCCACTGGATTCTTGTTTC
<i>windpipe</i>	TGGCAACCACAATGAGGAACAG	GACCGAGAAGACCTCCAGTCAAC
<i>Socs36E</i>	CAGTCAGCAATATGTTGTCG	ACTTGCAGCATCGTCGCTTC
<i>mef2</i>	ATCGGCAGGTGACCTTCAAC	GTTGTACTCGGTGTACTTGAGCAG

All primers listed 5' to 3'. Sequences from <sup>16</sup>, <sup>28</sup>, and FlyPrimerBank (<http://www.flyrnai.org/>).